system, since the liver plays an important role in endotoxicosis⁸ and endotoxin depletes carbohydrate reserves first of all from the liver4, and reverses gluconeogenesis; c) glucose feeding was prolonged until the 16th hour after endotoxin challenge, contrary to other studies cited above with only 1 or 2 injections and small quantities of glucose. Materials and methods. Swiss, male, albino mice (24 \pm 2 g) were housed on laboratory food and water ad libitum; food was withdrawn at the beginning of the experiments. To determine lethality, mice were injected with 500 ug of E. coli 026: B6 endotoxin (Difco, lot 586787) i.p. and the survival was recorded 48 h later. The LD_{50} was calculated according to Reed and Muench⁹. Glucose (5%), sodium chloride (0.9%), or water, in 0.5 ml, was given by stomach tube 1 h before and 1, 6, 11 and 16 h after endotoxin challenge.

Liver glycogen levels were determined according to the method of Kemp and Kits van Heijningen 10 as adapted previously 11. The glycogen level is expressed as mg/ 100 mg fresh liver weight. The results were evaluated by the Student t-test and the chi square test. A SE was calculated for all mean values.

Results and discussion. Data in the table show that the administration of a total of 125 mg glucose (5% solution)

Influence of glucose or saline feeding on endotoxin lethality in normal

Treatment	Living/Total (48 h)	Percent survival	Statistics
1. Water	9/25	36	
2. Physiological saline	11/27	40	2 VS 1 NS
3.5% glucose	10/25	40	3 VS 1 NS

NS = not significant.

by gavage (25 mg per dose, 1 h before and 1, 6, 11, 16 h after endotoxin challenge) did not alter endotoxin lethality. Oral administration of physiological saline was also without effect on the outcome of endotoxin lethality. There was 36%, 40.7% and 40% survival level in the water, physiological saline and 5% glucose treated groups, respectively.

Data in the figure clearly show that liver glycogen both in the water-treated and physiological glucose-treated groups was significantly lower within 4 h after endotoxin challenge, and continued so until a minimal carbohydrate level was achieved in both groups. The liver glycogen level did not increase in response to glucose feeding in endotoxin-challenged groups.

These results support earlier observations 4-6 obtained in endotoxin hyperreactive animals, with i.v. glucose administration, that exogenous glucose supply does not influence the outcome of endotoxin lethality. As in earlier studies 4,5 with parenteral glucose administration, exogenous glucose given by stomach tube also failed to prevent glycogen depletion during endotoxicosis (figure), either because it was metabolized and/or converted to muscle glycogen since mucsle glucogenesis is not impaired in endointoxicated animals4. It may be that liver glycogenesis is controlled at 2 levels; glucose-induced cycle can only proceed under normal conditions but cortisoneinduced phase can proceed just as well in endotoxin poisoned animals as in normal mice, although cortisonesparing of liver glycogen levels could only be secondary to protection against endotoxin lethality. Finally, the primary event in endotoxin death still remains elusive.

- M. K. Agarwal, Naturwissenschaften 62, 167 (1975).
- L. J. Reed and H. A. Muench, Am. J. Hyg. 27, 493 (1938).
- 10 A. Kemp and J. M. Kits van Heijningen, Biochem. J. 56, 646 (1954).
- 11 M. K. Agarwal, Biochem. Pharmac. 23, 2577 (1974).

Influence of anaesthesia by carbon dioxide and ether on locomotor activity in Drosophila melanogaster

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Summary. Locomotor activity of Drosophila melanogaster was investigated in 2 recently caught unrelated wild stocks. It is strongly affected by anaesthesia with either carbon dioxide or ether. Both anaesthetics had opposite effects. Ether produces a longlasting decrease in activity. Carbondioxide causes an increase of locomotor activity, but smaller and of shorter duration than the effect of ether.

Research on activity of Drosophila has shown that, it is a complex character, which is under control of both genetic and environmental factors 1-5. It could be expected that anaesthetics, used to immobilize flies, would affect the locomotor activity in later life. During experiments on locomotor activity, we obtained evidence that these effects could be considerable.

Therefore an experiment was started to investigate the long-term effects of both ether and CO₂ on this character. The locomotor activity of 2 wild strains of Drosophila melanogaster, recently collected in Dahomey (now Benin) and Spain (La Mancha) was measured in an apparatus that was especially constructed for this purpose. It is a modification of the apparatus, used by Ewing¹, and consists of a row of 20 vials, which can be transversed in one direction only.

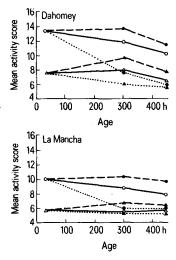
The mean activity score was calculated as the mean number of vials the flies have visited after 20 min. The activity tests were performed at 20 \pm 1 °C. Virgin flies were collected with either ether or CO2 and tested at ages of 30, 300 and 450 h. Some of the flies of the 300 and 450 h old flies were submitted to a subsequent anaesthesia with either CO2 or ether. The ether was tested on the presence of peroxides. The results were submitted to an analysis of variance. No interactions between the first

- A. W. Ewing, Anim. Behav. 11, 369 (1963).
- K. Connolly, Anim. Behav. 14, 444 (1966).A. Manning, Anim. Behav. 9, 82 (1961).
- B. Grant and L. E. Mettler, Genetics 62, 625 (1969).
- R. Hardeland and G. Stange, J. Insect Physiol. 17, 427 (1971).
- C. Montijn, F. R. Dijken, M. H. den Boer and W. Scharloo, D.I.S. 51, 151 (1974).

Mean activity scores of flies at ages of 30, 300 or 450 h (sexes pooled)

Age	1st anaesthetic	2nd anaesthetic	Dahomey	N	La Mancha	N
30	Ether		7.64	203	5.72	201
	CO ₂	_	13.29	186	9.92	199
300	Ether	Ether	6.09	196	5.29	181
	Ether	CO ₂	9.53	208	6.52	190
	Ether		7.57	201	5.32	191
	CO ₂	Ether	7.43	187	5.71	199
	CO ₂	CO ₂	13.48	188	10.15	184
	CO ₂	_	11.61	185	8.77	178
450	Ether	Ether	5.62	197	5.14	186
	Ether	CO ₂	7.70	184	6.00	182
	Ether		6.37	149	5.76	146
	CO ₂	Ether	5.86	188	5.80	181
	CO2	CO_2	11.23	186	9.56	187
	CO_2		10.11	149	7.86	174

N = Number of flies.



Effects of anaesthesia with CO2 and ether on locomotor activity of 2 stocks of Drosophila melanogaster. Circles represent flies collected with CO2, and triangles flies collected with ether. Solid symbols stand for subsequent anaesthesia. Dashed lines: subsequent anaesthesia with carbondioxide. Dotted lines: subsequent anaesthesia with ether.

- R. C. King and L. P. Wilson, J. exp. Zool. 130, 71 (1955). J. M. Perron, L. Huot, G. W. Corrivault and S. S. Chawla, J. Insect Physiol. 18, 1869 (1972).

and second treatment were found (p > 0.05). In the table and the figure we averaged the mean activity scores of males and females, because no significant differences were found between both sexes (p > 0.25). The most striking result from this experiment, which is highly significant (p < 0.001), is the overall activity-decreasing effect of ether anaesthesia throughout the experiment. The fact that ether has such a longlasting effect does not agree with the impression of many Drosophila-workers, who think that ether effects have disappeared after 24 h?. Contrary to ether, CO₂ causes an increase in activity, but this effect does not seem to last so long. Perron et al.8 found some toxic effects of CO2 in D. melanogaster on physiological characters but only when very young flies were treated over a long period.

In accordance with the results of Hardeland and Stange⁵, we did not detect age effects on the locomotor activity (p > 0.10). There is a difference in activity between the 2 stocks: flies of the Dahomey stock are more active than La Mancha flies (p < 0.01). This is probably correlated with the difference in body-size.9

The physiological bases of the anaesthetic effects are not known. King and Wilson 10 suggest that ether may upset the phosphorus turnover. Wigglesworth 11 proposed that oxygen lack caused by CO2-anaesthesia prevents the oxidation of acid metabolites produced by the activity of the insect. Hardeland and Stange⁵ pointed out that there could be a connection between locomotor activity and the activity of cytochrome-c-oxidase. We have shown that NADH-dehydrogenase activity in the mitochondria is highly correlated to locomotor activity (Thörig et al., unpublished, Scharloo et al., in press). Perhaps it can be supposed that the anaesthetics affect these enzymes by altering the structures of membranes 12.

This suggests the possibility of interference of the anaesthetics with the energy metabolism of the insect. It is clear that ether produces not only big, but also long, perhaps livelong effects on the locomotor activity in Drosophila melanogaster. Therefore it should be recommended to minimize the use of anaesthetics without careful analysis of their effect, especially when working on behavioural traits.

- R. Baptist and A. Robertson, Theoretical and applied genetics (1976).
- R. C. King and R. G. Burnet, D.I.S. 31, 130 (1957). 10
- C. R. Ribband, J. exp. Biol. 27, 302 (1950).
- 12 C. D. Richards, Nature 262, 534 (1976).

Effect of S-adenosyl-L-methionine¹ on ethynylestradiol-induced impairment of bile flow in female rats

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Summary. Prevention by S-adenosyl-L-methionine (SAMe) of the bile flow reduction induced by ethynylestradiol (EE) is demonstrated by comparing the flow rate and the bile salt concentration of bile in EE-treated animals with that in animals given both EE and SAMe.

Changes in bile flow and composition have been shown in the rat following EE-administration^{8,4}, and have been correlated with the impairment of the system responsible for the secretion of the bile salt-independent fraction of the canalicular bile 5. Moreover, it has been reported by various authors 6,7 that O-methylated derivatives of 2hydroxyestrogens constitute the major fraction of urinary estrogens.

- SAMe was supplied by BioResearch Co., 20060 Liscate (Milano), Italy.
- Clinica Medica III, University of Milan, Italy.
- T. A. J. Heikel and G. H. Lathe, Br. J. Pharmac. 38, 593 (1970).
- R. A. Davis and F. Kern, Gastroenterology 70, 1130 (1976). J. J. Gumucio and V. D. Valdivieso, Gastroenterology 61, 339 (1971).
- J. Fishman, J. clin. Endocr. Metab. 23, 207 (1963).
- P. Ball, H. P. Gelbke and R. Knuppen, J.clin. Endocr. Metab. 40, 406 (1975).